

STEVEN HARDWICK

Address: Department of Biochemistry, University of Cambridge, 80 Tennis Court Road, Cambridge, UK, CB2 1GA.
Phone: Work: +44(0)1223-766020
Email: swh32@cam.ac.uk

Education and Research Experience

2007- **Research Associate**
Department of Biochemistry, University of Cambridge
Group Leader: Prof. B. F. Luisi

Synopsis: The main focus of research in our group is on a bacterial multi-protein RNA processing machine termed the RNA-degradosome. The full degradosome from *Escherichia coli*, and the individual protein components of the complex, Endoribonuclease E (RNase E), a DEAD-box RNA helicase (RhlB), the glycolytic enzyme Enolase and the phosphorolytic exoribonuclease PNPase have been studied by many biochemical and biophysical techniques including isothermal titration calorimetry, surface plasmon resonance, and primarily X-ray crystallography. I have been primarily studying an RNA degradosome complex in *Caulobacter crescentus*, and have identified by co-immunoprecipitation a complex composed minimally of RNase E, a DEAD-box RNA helicase similar to the *E. coli* RhlB, PNPase and the TCA cycle enzyme Aconitase. I have also solved the crystal structure of the *Caulobacter crescentus* PNPase bound to endogenous RNA, revealing asymmetry and potential allostery in the mechanism of RNA binding and processing. The current focus of my research is to use *C. crescentus* as a model of bacterial cell development, and determine whether the degradosome complex is the target of cell cycle dependent regulation.

I am additionally involved with several other projects within the lab studying various proteins involved in regulating RNA metabolism in bacteria. These include the RNA chaperone/binding proteins ProQ and RapZ from *E. coli*, and a bacteriophage encoded inhibitor of RNase E we have termed Dip (Degradosome interacting protein).

2003-2007 **PhD in Structural Biology – Newcastle University, UK.**
Title: Structural studies of regulators of sigma B in gram positive bacteria.
Supervisor: Prof. R. J. Lewis

Synopsis: When the bacteria *B. subtilis* and *S. aureus* encounter environmental or physical stresses the alternative sigma factor, σ^B directs RNA polymerase to the promoters of over 200 genes encoding the general stress proteins. The activity of σ^B is controlled by a set of proteins termed regulators of sigma B (Rsb).

Throughout my PhD I studied some of the interactions between the Rsb proteins, predominantly the activation of the type 2C protein phosphatase RsbU via an interaction with its activator protein RsbT.

The *B. subtilis* protein RsbU is composed of a C-terminal type 2C phosphatase domain and an N-terminal domain presumed to be involved in binding to the phosphatase activator protein RsbT, an interaction that leads to a 20 fold increase in phosphatase activity towards phosphorylated RsbV (2). I determined experimentally that the N-terminal domain of RsbU is indeed an RsbT binding domain, and furthermore I discovered four residues on the surface of N-RsbU crucial for the binding to RsbT.

2000-2003 Newcastle University, UK.
BSc Hons. Medical Microbiology (Grade 2.1)
Including research project: Identification of the cadmium binding site of CtpR in Mycobacteria in vivo
Supervisor: Dr. J. S. Cavet.

Teaching Experience:

2014-2017 Session chair – PartIII research symposium
Lead questions/discussion during “Structural Biology and Drug Discovery” session at Masters students research presentation session

2013- Honorary Post-doctoral teaching fellow
Participant in 2nd and 3rd year undergraduate group supervisions
Demonstrator in undergraduate bio-informatics practical sessions
Lead small group supervisions on data handling exercises and journal club research projects for 2nd and 3rd year undergraduate students

2007- Supervisor for final year undergraduate research projects
Introduced project background, explained theory behind experimental approaches, demonstrated laboratory techniques, provided constructive feedback on project reports

Collaborations

2016- Dr Jared Schrader (Wayne State University – Detroit USA)
Recently established collaboration to study the RNA degradosome in *Caulobacter crescentus*

2014- Prof. Rob Lavigne (KU Leuven, Belgium)
Collaboration to study the structure and function of the phiKZ phage encoded inhibitor of the bacterial RNA degradosome

2012- Prof Marilis Marques (University of Sao Paulo, Brazil)
Ongoing collaboration to study role of RNA helicases in *Caulobacter crescentus*
Joint FAPESP funded travel grant applied for to fund collaboration (2014)

2012- Prof Hagan Bayley (University of Oxford, UK)
Collaboration established to examine possibility of using Polynucleotide phosphorylase to aid nano-pore sequencing of RNA
Co-authored publication

2011- Prof Urs Jenal (Biozentrum, Universitat Basel, Switzerland)
Collaboration initiated to study in vivo role of RNA degradosome in *Caulobacter crescentus*
Co-authored publication

Peer Review Experience

I have co-reviewed research papers and reviews submitted to:
Nature, Molecular Cell, Cell Reports, Structure, PLOS Genetics, Biochemistry, Journal of Biological Chemistry, BBA gene regulatory mechanisms, FEBS Journal, Molecular Microbiology, RNA journal, Nucleic Acids Research, Journal of Molecular Microbiology and Biotechnology, Acta Crystallographica section F

Organisational and Administrative Experience

2014 Deputy X-ray crystallography facility manager

2008- Assisted with lab consumables ordering/purchasing

Personal Development

- 2013- Attended workshops on effective presentation skills as part of Post-doctoral teaching fellows scheme (University of Cambridge)
- 2013 Workshop for supervising undergraduates (Personnel Dept, University of Cambridge)
- 2009 British Council organised workshop "Establishing a successful international research career" (Lisbon, Portugal)

Invited Presentations and Conference Attendance

- 2016 81st Harden Conference: RNA and Disease (Winchester, UK). Talk presented "A novel mechanism for bacteriophage encoded inhibition of the RNA degradosome assembly"
- 2014 EMBO Workshop: "Stalked alpha-proteobacteria and their relatives" (Marburg, Germany). Talk presented "Molecular Recognition through micro-domains in the *Caulobacter crescentus* RNA degradosome"
- 2013 "Regulating with RNA in Bacteria" (Wurzburg, Germany). Poster presented "Characterisation of the RNA degradosome in *Caulobacter crescentus*."
- 2012 FASEB meeting "Post-transcriptional control of gene expression: mechanisms of mRNA decay" (Steamboat Springs, Colorado, USA). Talk presented "The RNA degradosome of *Caulobacter crescentus* and its dynamic association with 3'-5' exoribonucleases"
- 2011 Diamond synchrotron user meeting (Didcot UK). Invited speaker "Improved diffraction of *C. crescentus* PNPase crystals by dehydration with HC1."
- 2011 Mol Micro meeting Wurzburg (Wurzburg, Germany). Poster presented "The crystal structure of *Caulobacter crescentus* polynucleotide phosphorylase reveals a potential mechanism for RNA substrate channelling"
- 2009 3rd ASM conference on Prokaryotic development (Cambridge, Massachusetts, USA). Poster presented "A multi-enzyme RNA degradative machine from *Caulobacter crescentus*."
- 2008 FASEB meeting "Post-transcriptional control of gene expression: mechanisms of mRNA decay" (Lucca, Italy). Talk presented "Identification of an RNA degradosome complex in *Caulobacter crescentus* that is regulated in a cell cycle dependent manner"

Original Research Publications

- 2017 Tichy EM, **Hardwick SW**, Luisi BF, Salmond GPC. 1.8 Å resolution crystal structure of the carbapenem intrinsic resistance protein CarF. *Acta Cryst D*. **73**, 549-556
- 2017 Aguirre AA, Vicente AM, **Hardwick SW**, Alvelos DM, Mazzon RR, Luisi BF, Marques MV. Association of the Cold Shock DEAD-Box RNA Helicase RhIE to the RNA Degradosome in *Caulobacter crescentus*. *J. Bacteriol.* **199**, e00135-17
- 2017 Gonzalez GM, **Hardwick SW**, Maslen SL, Skehel JM, Holmqvist E, Vogel J, Bateman A, Luisi BF, Broadhurst RW. Structure of the *Escherichia coli* ProQ RNA-binding protein. *RNA* **23**, 696-711

- 2016** Van den Bossche A, **Hardwick SW**, Ceyskens PJ, Hendrix H, Voet M, Dendooven T, Bandyra KJ, De Maeyer M, Aertsen A, Noben JP, Luisi BF, Lavigne R. Structural elucidation of a novel mechanism for the bacteriophage-based inhibition of the RNA degradosome. *Elife* **5**, e16413
- 2014** Voss J, Luisi BF, **Hardwick SW**. Molecular recognition through microdomains in the *Caulobacter crescentus* RNA degradosome. *Nucleic Acids Res.* **42**, 13294-305
- 2013** Ayub M, **Hardwick SW**, Luisi BF, Bayley H. Nanopore-based identification of individual nucleotides for direct RNA sequencing. *Nano Lett.* **13**, 6144-50
- 2013** Pietras Z, **Hardwick SW**, Swiezewski S, Luisi BF. Potential regulatory interactions of *Escherichia coli* RraA protein with DEAD-box helicases. *J Biol Chem.* **288**, 31919-29
- 2012** **Hardwick SW**, Gubbey T, Hug I, Jenal U, Luisi BF. Crystal structure of *Caulobacter crescentus* polynucleotide phosphorylase reveals a mechanism of RNA substrate channeling and RNA degradosome assembly. *Open Biology* 2012 2:120028
- 2011** **Hardwick SW**, Chan VS, Broadhurst RW, Luisi BF. An RNA degradosome assembly in *Caulobacter crescentus*. *Nucleic Acids Res.* **39**, 1449-59
- 2009** Pané-Farré J, Jonas B, **Hardwick SW**, Gronau K, Lewis RJ, Hecker M, Engelmann S. Role of RsbU in controlling SigB activity in *Staphylococcus aureus* following alkaline stress. *J Bacteriol.* **191**, 2561-73
- 2007** **Hardwick SW**, Pané-Farré J, Delumeau O, Marles-Wright J, Murray JW, Hecker M, Lewis RJ. Structural and functional characterization of partner switching regulating the environmental stress response in *Bacillus subtilis*. *J Biol Chem.* **282**, 11562-72
- 2004** Delumeau O, Dutta S, Brigulla M, Kuhnke G, **Hardwick SW**, Völker U, Yudkin MD, Lewis RJ. Functional and structural characterization of RsbU, a stress signaling protein phosphatase 2C. *J Biol Chem.* **279**, 40927-37

Reviews/Points of View

- 2017** Dendooven T, Van den Bossche A, Hendrix H, Ceyskens PJ, Voet M, Bandyra KJ, De Maeyer M, Aertsen A, Noben JP, **Hardwick SW**, Luisi BF, Lavigne R. Viral interference of the bacterial RNA metabolism machinery. *RNA Biol.* **14**, 6-10
- 2013** **Hardwick SW**, Luisi BF. Rarely at rest: RNA helicases and their busy contributions to RNA degradation, regulation and quality control. *RNA Biol.* **10**, 56-70

Academic Referees

Professor Ben Luisi, Department of Biochemistry, University of Cambridge, 80 Tennis Court Road, CB2 1GA, UK. 01223-766019, bfl20@cam.ac.uk

Professor Rick Lewis, Institute of Cell and Molecular Biosciences, Newcastle University, UK. 01912085482, rick.lewis@ncl.ac.uk